

# AmplifyRP® Acceler8® for Little cherry virus 2 (LChV2)

## Rapid RNA Test Kit

Product No. ACS 85601/0008



### Intended Use:

AmplifyRP Acceler8 for LChV2 is a rapid RNA amplification and detection platform designed for field-based or laboratory testing of cherry crops for *Little cherry virus 2*.

This test has been validated to detect isolates from the USA, Canada, and Australia including USA6b, LC5, and C14.

This test does not cross-react with *Cherry leaf roll virus (CLR)*, *Tomato bushy stunt virus (TBSV)*, *Plum pox virus (PPV)*, *Western X-disease phytoplasma (WX)*, or *Little cherry virus 1 (LChV1)*.

### Kit Storage:

Kit components should be stored refrigerated (2 - 8 °C), unless specified otherwise below:

**\*\*\* Reaction pellets should be stored at -10 to -30 °C when not in use. \*\*\***

Before use, allow all kit components to warm to room temperature (18 - 30 °C) for 10 to 20 minutes.

### Sample Preparation

*NOTE: AmplifyRP is a very sensitive molecular assay. Do not re-use disposable kit components. It is recommended that latex gloves be worn when taking samples and performing assay. If wearing latex gloves, change them between samples and test runs. Sanitize work area and non-disposable equipment between runs with a 10 % bleach solution.*

1. LChV2 in infected cherry trees can be unevenly distributed and variable in virus titer throughout a year. These factors can limit the ability of detecting the virus. Collect samples from symptomatic limbs, otherwise collect samples from throughout the tree canopy. LChV2 concentration is higher in one-year old limbs, so sample leaves from established limbs or at the base of the current year's growth. Testing leaves in late spring to mid-summer, or dormant budwood increases the chance of detecting the virus.
2. Cut or tear the basal portion from 3 to 5 leaves including portions of the attached petioles (Figure 1) or collect sections of budwood bark. The total sample weight should be approximately 0.3 g. To avoid contamination, make certain you clean cutting instruments between samples with a 10 % bleach solution.
3. Insert the sample between the mesh linings of the GEB3 sample extraction bag. The sample can be extracted by rubbing the outside of the bag with a blunt object such as a pen or marker on a hard surface (Figure 2). Once the sample has been thoroughly homogenized, it is ready to be tested.

### Amplification

1. Allow heat block to warm to 39 °C before preparing reactions. If using an Agdia supplied heat block, allow 2 to 3 minutes for this step.
2. Remove the strip of reaction pellets from the desiccated container included in the kit. While securing the strip of pellets in a 200 µL PCR tube rack, cut the number of reaction pellets from the strip that are intended for use. Immediately place remaining reaction pellets back into the desiccated tube for later use.
3. Dispense 10 µL of PD1 Pellet Diluent into a reaction pellet. Be sure the pellet is visible in the bottom of the tube before opening or dispensing liquid into the tube.
4. Using a 1 µL loop or pipette, immediately transfer 1 µL of sample extract into the rehydrated reaction pellet. Recap the tube and mix by flicking the bottom of the tube 6 to 8 times or by using a laboratory vortex mixer. Then shake or centrifuge the liquid contents to the bottom of the tube.
5. Add reaction to the portable heat block for 20 minutes.
6. Immediately remove reaction from heat block and proceed to detection steps.

### Contents of Kit:

- Reaction pellets (8)
- Amplicon detection chambers (8)
- PD1 pellet diluent (1.0 mL)
- GEB3 sample extraction bags (8)
- 1 µL transfer loops (10)

### Not Included but Required:

- Portable heat block (ACC 00592)
- 10 µL pipette (ACC 00770/0010)
- 10 µL pipette tips (ACC 00160)
- 200 µL PCR tube rack (ACC 00170)

\* A starter pack inclusive of the items not included above can be purchased from Agdia (ACC 00150).

Figure 1. LChV2 Infected Leaf

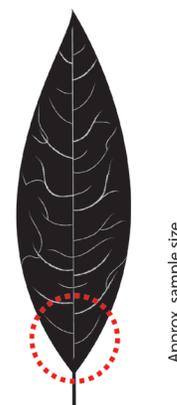


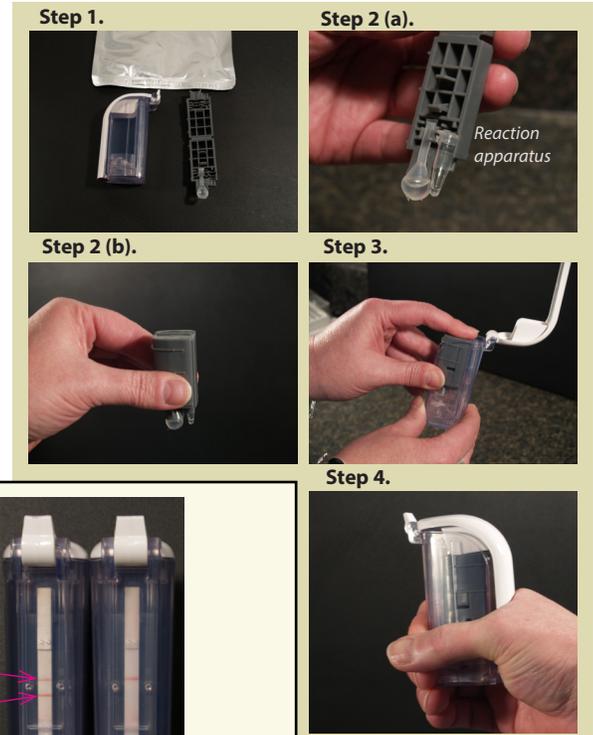
Figure 2. Sample Extraction



## Detection

In order to avoid possible contamination of future tests, DO NOT open the reaction pellet.

1. Open the foil pouch containing the Amplicon detection chamber. There are two pieces to the chamber as indicated in the figure to the right.
2. a.) Add the unopened reaction tube to reaction apparatus as illustrated to the right. b.) Once the tube has been added, snap the apparatus shut which will immobilize the reaction tube.
3. Add the reaction apparatus to the detection chamber housing as indicated. **IMPORTANT: The reaction tube should be facing toward the lateral flow strip, contained in the housing, during this step.**
4. Push down on the handle of the detection chamber housing until it snaps shut. Wait 20 minutes before interpreting results. Positive results may be visible in as little as 5 to 10 minutes. Samples that contain lower copy numbers may take up to 20 minutes to produce a positive test line.



## Interpret Results

Result	ImmunoStrip Reaction
<b>Positive</b>	Control and Test lines are both visible.
<b>Negative</b>	Control line is visible. Test line not visible.
<b>Invalid</b>	Control line not visible.



## Limitations

The following is a description of factors that could limit test performance or interfere with proper test results.

**Sample Extraction Buffer:** This test must be used with the supplied sample extraction buffer to obtain optimal results.

**Addition of sample extract to reaction pellet:** It is important to add only the prescribed amount of sample extract to reaction pellets. Adding too much extract may cause test failure.

**Storage:** Test results may be weak or the test may fail if the storage instructions are not followed properly. The lyophilized test components must remain protected from light to prevent bleaching and sealed with desiccant when not in use to prevent moisture degradation, which may affect test results. Do not store pellets at temperatures greater than 42 °C, as this may cause test failure.

### Questions or Technical Support:

Phone: 800-622-4342 (toll-free) or 574-264-2014

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E-mail: [amplifyrp@agdia.com](mailto:amplifyrp@agdia.com) for general questions about this assay or AmplifyRP technology  
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